

# Effect of menbutone on oxidative stress in sheep

## Efecto de la menbutona sobre el estrés oxidativo en ovejas

Rahmi Canbar<sup>1</sup>, Tugba Melike Parlak<sup>2\*</sup>, Muhittin Uslu<sup>3</sup>, Enver Yazar<sup>2</sup>

<sup>1</sup>Aksaray University, Faculty of Veterinary Medicine, Department of Pharmacology and Toxicology. Aksaray, Türkiye.

<sup>2</sup>Selcuk University, Faculty of Veterinary Medicine, Department of Pharmacology and Toxicology. Konya, Türkiye.

<sup>3</sup>Yozgat Bozok University, Sefaattü Vocational School, Veterinary Laboratory and Veterinary Health Department. Yozgat, Türkiye.

\*Corresponding author: [tugba.parlak@selcuk.edu.tr](mailto:tugba.parlak@selcuk.edu.tr)

### ABSTRACT

The objective of this research was to determine the effects of administering the choleric-effective menbutone (genabilic acid) at the recommended dose and duration of serum oxidative status and biochemical parameter levels in sheep. As is known, changes in the oxidative status of living organisms can be observed in many drug applications, disease states, or metabolic disorders. In the study, menbutone was administered at a dose of 10 mg·kg<sup>-1</sup> (slow IV, SID, 2 days) to 10 Central Anatolian Merino sheep. Blood samples were collected from the sheep before (0<sup>th</sup> hour) and at the 12<sup>th</sup>, 24<sup>th</sup>, and 48<sup>th</sup> hours after menbutone administration. Serums were obtained from the collected blood samples. Serum oxidative status parameters (8-hydroxy-2-deoxyguanosine, malondialdehyde, superoxide dismutase, glutathione peroxidase, catalase), troponin I, and creatine kinase-MB isoenzyme levels were measured using the ELISA method, while levels of serum routine biochemical parameters were measured using an autoanalyzer. Menbutone did not affect oxidative stress parameters (8-hydroxy-2-deoxyguanosine, malondialdehyde, superoxide dismutase, glutathione peroxidase, catalase) in sheep ( $P>0.05$ ) but caused temporary increases ( $P<0.05$ ) in serum triglyceride and cholesterol levels. However, no statistically significant changes were detected in other serum biochemical parameters (Troponin I, creatine kinase-MB isoenzyme, alkaline phosphatase, aspartate aminotransferase, alanine aminotransferase, low-density lipoprotein, high-density lipoprotein, blood urea nitrogen, creatinine). In conclusion, it can be asserted that menbutone can be used safely in sheep without altering oxidative stress parameters, but monitoring serum triglycerides and cholesterol may be important in animals with metabolic disorders.

**Key words:** Menbutone; oxidative status; biochemical parameters; sheep

### RESUMEN

El objetivo de esta investigación fue determinar los efectos de la administración de menbutona (ácido genabilico), con efecto colerético, en la dosis recomendada y la duración del estado oxidativo sérico y los niveles de parámetros bioquímicos en ovejas. Como se sabe, los cambios en el estado oxidativo de los organismos vivos se pueden observar en muchas aplicaciones de fármacos, enfermedades o trastornos metabólicos. En el estudio, se administró menbutona en una dosis de 10 mg·kg<sup>-1</sup> (IV lenta, SID, 2 días) a 10 ovejas merinas de Anatolia central. Se recogieron muestras de sangre de las ovejas antes (hora 0) y a las 12, 24 y 48 horas después de la administración de menbutona. Se obtuvieron sueros de las muestras de sangre recogidas. Los parámetros séricos del estado oxidativo (8-hidroxi-2-desoxiguanosina, malondialdehído, superóxido dismutasa, glutatión peroxidasa, catalasa), troponina I y los niveles de isoenzima MB de la creatina quinasa se midieron mediante el método ELISA, mientras que los niveles de parámetros bioquímicos séricos de rutina se midieron utilizando un autoanализador. La menbutona no afectó los parámetros de estrés oxidativo (8-hidroxi-2-desoxiguanosina, malondialdehído, superóxido dismutasa, glutatión peroxidasa, catalasa) en ovejas ( $P>0,05$ ), pero causó aumentos temporales ( $P<0,05$ ) en los niveles séricos de triglicéridos y colesterol. Sin embargo, no se detectaron cambios estadísticamente significativos en otros parámetros bioquímicos séricos (troponina I, isoenzima MB de la creatina quinasa, fosfatasa alcalina, aspartato aminotransferasa, alanina aminotransferasa, lipoproteínas de baja densidad, lipoproteínas de alta densidad, nitrógeno ureico en sangre, creatinina). En conclusión, se puede afirmar que la menbutona puede usarse de forma segura en ovejas sin alterar los parámetros de estrés oxidativo, pero la monitorización de los triglicéridos y el colesterol séricos puede ser importante en animales con trastornos metabólicos.

**Palabras clave:** Menbutona; estado oxidativo; parámetros bioquímicos; ovejas

## INTRODUCTION

Menbutone (genablic acid) is a drug licensed for use in the Veterinary field for the treatment of metabolic diseases related to the digestive system and liver in horses (*Eqqus caballus*), cattle (*Bos taurus*), sheep (*Ovis aries*), goats (*Capra hircus*), and pigs (*Sus scrofa domesticus*) for choleric purposes. Although the mechanism of action of the drug is not clearly known, it has been reported that it may exert its effect by stimulating the parasympathetic nervous system [1, 2].

The general dosage of the drug in target species is 10 mg·kg<sup>-1</sup> administered intramuscularly (IM) or intravenously (IV). Menbutone causes a 2 to 5-fold increase in bile, liver, stomach, and pancreatic secretions after its administration. It directly stimulates liver parenchymal cells, initiating the secretion of bile in its normal composition and in abundant quantities. This improves digestive and liver function in patients [2, 3].

The use of the drug is not recommended in patients with heart disease and in the late stages of pregnancy. Drug may cause some adverse effects such as restlessness, salivation, lacrimation, tremors, involuntary defecation and urination, inability to stand during rapid intravenous administration, as well as necrosis, edema, and hemorrhage at the injection site in target species. When administered at high doses to rodents, it has been determined that it may cause cyanosis, respiratory depression, nephrotoxic and hepatotoxic effects [1, 3].

Menbutone increases bile secretion in the liver [1], and it has been reported bilirubin exerts an antioxidant effect by scavenging or neutralizing free radicals. Bile acids, produced in the liver, can indirectly exert an antioxidant effect by facilitating the absorption of vitamins such as A and E from the intestines. Furthermore, bile acids can exhibit antioxidant effects by trapping oxygen free radicals within bile acid micelles [4]. Free oxygen radicals are the most produced substances in living organisms, and these harmful structures are converted into harmless structures by enzymatic (superoxide dismutase, catalase, glutathione peroxidase, enter other) [5], and/or non-enzymatic substances (vitamin C, vitamin E, vitamin A, glutathione, etc.) [6].

Specifically, an oxygen molecule that accepts an electron during the mitochondrial respiratory chain forms the superoxide radical. The superoxide dismutase (SOD) enzyme found in cells combines two superoxide radicals with two hydrogen atoms to form hydrogen peroxide. Then hydrogen peroxide is transformed into molecular oxygen and water by the enzymes glutathione peroxidase (GPX) and catalase (CAT), turning it into harmless molecules [5].

There is an equilibrium between antioxidants and oxidants in living organisms. A disruption of this balance in favor of oxidant substances is defined as oxidative stress. Non-neutralized free oxygen radicals attack structures in cells such as lipids, proteins, and DNA, causing damage to these structures. Lipids in the cell membrane are highly sensitive to free oxygen radicals, and damage to this structure is defined as lipid peroxidation [5]. Lipid peroxidation produces malondialdehyde (MDA) and determination of MDA level is used to understand the degree of cellular damage [7].

Malondialdehyde has mutagenic, cytotoxic, and carcinogenic properties. MDA can also inhibit the functions of antioxidant enzymes associated with protecting cells against oxidative stress [8]. 8-hydroxy-2-deoxyguanosine (8-OHdG), produced during the repair of DNA injury caused by oxidative stress, is considered as the most common biomarker of DNA damage [9] and has mutagenic potential [10].

It has been suggested that some drugs causing oxidative stress may cause heart damage and that markers of heart damage, such as troponin I and creatine kinase-MB isoenzyme (CK-MBiso) in serum, may increase [11, 12, 13].

Although sheep are defined as the target species in the use of menbutone [14], there is very limited number of studies on its effect on organ damage biomarkers in sheep. It is known that drugs used in treatment may cause some undesirable or side effects at the recommended doses, in addition to their therapeutic effects in living organisms. Drug-induced undesirable or side effects can be clinically observed and can also be identified by measuring certain parameters in the blood [15].

The levels of these analyzed parameters allow us to understand the damage developing in the systems or organs of living organisms [15, 16]. CK-MB isoenzyme and troponin I levels are associated with heart damage [17, 18], while creatinine and blood urea nitrogen (BUN) levels provide information about kidney function [19].

While serum alkaline phosphatase (ALP), aspartate aminotransferase (AST) and alanine aminotransferase (ALT) levels provide information about liver and bile duct function [20], triglyceride, cholesterol, low-density lipoprotein (LDL), and high-density lipoprotein (HDL) levels provide information about lipid metabolism [21].

Given that menbutone, which has a choleric effect, increases bile secretion [1, 14], may affect the oxidative mechanisms of bile content [4] and may cause serum biochemical values at recommended therapeutic doses and duration, it was hypothesized that the administration of menbutone to sheep may affect serum oxidative status and biochemical values.

The objective of this study was to determine the effect of menbutone administration in sheep on oxidative stress markers (8-OHdG, MDA, SOD, GPX, and CAT), cardiac (troponin I, CK-MBiso), hepatic (ALP, ALT, AST), and renal (BUN, creatinine) functions, alongside lipid profile parameters (cholesterol, triglycerides, HDL, LDL). Hence, this study will determine the drug's safety in sheep, in addition to its effect on the oxidative status.

## MATERIALS AND METHODS

### Experimental design and animal applications

The study was conducted on 10 Central Anatolian Merino sheep (2–3 years old, 50–55 kg). The procedure of the study was approved by the Ethics Committee of the Selcuk University Faculty of Veterinary Medicine Experimental Animal Production and Research Center (SUVDAMEK) (2025/63).

Menbutone (Bavet Menbutone inj., Bavet Ilac Sanayi, Istanbul, Türkiye) was administered to the sheep at a dose of 10 mg·kg<sup>-1</sup>

[slow IV, once daily (SID)] for 2 days (d). To minimize variations due to individual differences, no separate control group was established in the study. Instead, blood was collected from all animals 0 hours (h) before treatment and assessed as a self-control group.

Blood samples were taken from the jugular veins of the sheep before administration (0<sup>th</sup> h) and at 12<sup>th</sup>, 24<sup>th</sup>, and 48<sup>th</sup> hours after the first administration. To isolate serum from the collected blood samples, they were centrifugated (Sigma 3K18, Osterode am Harz, Germany) at 4000 g for 10 minutes and kept at – 80 °C (Operon Co Ltd Ultra Low Temperature, South Korea) until biochemical measurements.

### ELISA and blood serum biochemical analyses

Commercial ELISA kits (BT-LAB, Zhejiang, China) were used in the study, and all kits were selected to be specific to sheep. Serum 8-OHDG, MDA, SOD, GPX, CAT, troponin I, and CK-MBiso levels were measured using ELISA kits and an ELISA plate reader (MWGt Lambda Scan 200; Bio-Tec Instruments, Winooski, VT, USA) according to the kit instructions.

Serum AST, ALT, ALP, cholesterol, triglycerides, HDL, LDL, BUN, and creatinine levels were determined using an autoanalyzer (BT-3000 plus, Biotechnica Instruments, Rome, Italy).

### Statistical analysis

Research results are given as mean  $\pm$  standard error (SE) and were compared amongst groups and time points using ANOVA and post-hoc TUKEY tests (SPSS 29.0). The value of  $P < 0.05$  was accepted as statistically significance level.

## RESULTS AND DISCUSSION

TABLE I shows the effect of menbutone administration on oxidative stress parameters (SOD, GPX, CAT, MDA, 8-OHDG) in sheep, while TABLE II shows its effects on biochemical parameters (Troponin I, CK-MBiso, AST, ALT, ALP, cholesterol, triglyceride, HDL, LDL, BUN, creatinine). Menbutone administration had no effect on oxidative stress parameters in sheep ( $P > 0.05$ ). Menbutone caused a temporary increase in triglyceride and cholesterol levels in sheep ( $P < 0.05$ ), but no effect on heart, liver, and kidney function parameters was detected ( $P > 0.05$ ). Furthermore, no clinically adverse effects (fever, salivation, convulsions, decreased appetite, discomfort, etc.) were observed in the animals during the application period.

Menbutone is used in sheep for choleric purposes in the treatment of metabolic diseases related to the digestive system and liver. The drug causes an increase in bile, liver, stomach, and pancreatic secretions, exerting a regulatory effect on digestive and liver functions. General dosage of the menbutone in sheep is 10 mg·kg<sup>-1</sup> (IM or IV) [2, 3].

However, menbutone may restlessness, salivation, lacrimation, tremors, involuntary defecation and urination, etc. as side effects [1, 2, 3]. Although the drug has been used for a long time in target species and the bile acids and bilirubin it causes to be secreted are known to exhibit antioxidant effects, there are insufficient reports on its effects on oxidative status values and heart, liver, and kidney

**TABLE I**  
The effect of menbutone (10 mg·kg<sup>-1</sup>, slow intravenously, single in a day, 2 days) on serum oxidative status parameters in ten sheep (Mean  $\pm$  Standard error)\*

Parameters	0 <sup>th</sup> hour	12 <sup>th</sup> hour	24 <sup>th</sup> hour	48 <sup>th</sup> hour
SOD ng·mL <sup>-1</sup>	64.63 $\pm$ 5.25	63.10 $\pm$ 4.99	75.17 $\pm$ 7.02	66.14 $\pm$ 4.75
GPX nU·mL <sup>-1</sup>	606.48 $\pm$ 118.05	483.81 $\pm$ 79.50	544.81 $\pm$ 94.98	434.77 $\pm$ 67.33
CAT ng·mL <sup>-1</sup>	3.17 $\pm$ 0.60	3.61 $\pm$ 0.57	3.63 $\pm$ 0.64	3.24 $\pm$ 0.62
MDA nmol·L <sup>-1</sup>	5.09 $\pm$ 0.82	4.18 $\pm$ 0.66	5.52 $\pm$ 1.16	3.79 $\pm$ 0.55
8-OHDG ng·mL <sup>-1</sup>	5.53 $\pm$ 0.95	4.84 $\pm$ 0.58	5.58 $\pm$ 0.75	4.65 $\pm$ 0.68

SOD: Superoxide dismutase, GPX: Glutathione peroxidase, CAT: Catalase, MDA: Malondialdehyde, 8-OHDG: 8-hydroxy-2-deoxyguanosine. \*: No statistical difference was found in the data in the same row (ANOVA, Tukey test,  $P > 0.05$ )

**TABLE II**  
The effect of menbutone (10 mg·kg<sup>-1</sup>, slow intravenously, single in a day, 2 days) on serum biochemistry parameters in ten sheep (Mean  $\pm$  Standard error)

Parameters	0 <sup>th</sup> hour	12 <sup>th</sup> hour	24 <sup>th</sup> hour	48 <sup>th</sup> hour
Troponin I ng·mL <sup>-1</sup>	121.81 $\pm$ 27.81	192.81 $\pm$ 34.80	139.43 $\pm$ 31.04	118.97 $\pm$ 16.43
CK-MBiso ng·mL <sup>-1</sup>	0.67 $\pm$ 0.11	0.67 $\pm$ 0.08	0.63 $\pm$ 0.09	0.58 $\pm$ 0.07
ALP U·L <sup>-1</sup>	49.50 $\pm$ 15.17	101.70 $\pm$ 38.15	48.90 $\pm$ 16.33	74.00 $\pm$ 31.15
AST U·L <sup>-1</sup>	97.90 $\pm$ 4.19	100.50 $\pm$ 4.05	94.80 $\pm$ 4.51	94.70 $\pm$ 4.32
ALT U·L <sup>-1</sup>	187.40 $\pm$ 33.10	174.50 $\pm$ 36.26	193.20 $\pm$ 34.93	172.80 $\pm$ 38.44
Cholesterol mg·dL <sup>-1</sup>	67.20 $\pm$ 2.85 <sup>ab</sup>	73.40 $\pm$ 2.14 <sup>a</sup>	64.80 $\pm$ 2.71 <sup>ab</sup>	61.50 $\pm$ 3.59 <sup>b</sup>
Triglyceride mg·dL <sup>-1</sup>	16.10 $\pm$ 1.79 <sup>b</sup>	38.40 $\pm$ 2.36 <sup>a</sup>	19.70 $\pm$ 2.34 <sup>b</sup>	17.40 $\pm$ 2.04 <sup>b</sup>
LDL mg·dL <sup>-1</sup>	13.50 $\pm$ 0.80	13.00 $\pm$ 1.09	11.30 $\pm$ 0.91	11.30 $\pm$ 1.21
HDL mg·dL <sup>-1</sup>	32.80 $\pm$ 1.20	33.71 $\pm$ 1.62	31.99 $\pm$ 1.62	31.20 $\pm$ 1.75
BUN mg·dL <sup>-1</sup>	12.18 $\pm$ 0.41	10.48 $\pm$ 0.35	10.65 $\pm$ 0.69	12.40 $\pm$ 0.53
Creatinine mg·dL <sup>-1</sup>	0.91 $\pm$ 0.04	0.97 $\pm$ 0.04	0.88 $\pm$ 0.03	0.91 $\pm$ 0.03

CK-MBiso: Creatine kinase-MB isoenzyme, ALP: Alkaline phosphatase, AST: Aspartate aminotransferase, ALT: Alanine aminotransferase, LDL: Low-density lipoprotein, HDL: High-density lipoprotein, BUN: Blood urea nitrogen. <sup>a,b</sup>: Different letters in the same row are statistically significant (ANOVA, Tukey test,  $P < 0.05$ )

function parameters in healthy sheep. Furthermore, there is limited data on its direct effects on lipid metabolism parameters in sheep.

The effect of menbutone on some oxidative stress serum parameters in sheep were investigated in sheep which is target species. It was determined that administering menbutone at a dose of 10 mg·kg<sup>-1</sup> did not alter SOD, GPX, CAT, MDA, and 8-OHDG levels in sheep ( $P > 0.05$ , TABLE I). Following menbuton administration, it is expected that the increased secretion of antioxidant bile acids and bilirubin will also exhibit antioxidant effects. Hence, menbuton may reduce oxidative status parameters, even indirectly. However, there is no literature on the effects of menbutone on oxidative stress parameters. Studies on the effects of drugs on oxidative stress parameters in sheep frequently examine serum levels of SOD, GPX, CAT, MDA, and 8-OHDG [13, 22, 23, 24].

It is stated that the SOD enzyme is responsible for converting two superoxide radicals into hydrogen peroxide by combining them with two hydrogen atoms. It is reported that the GPX and

CAT enzymes inactivate hydrogen peroxide by converting it into molecular oxygen and water [5]. MDA is formed as a result of lipid peroxidation and is accepted as a biomarker of oxidative stress [7].

Furthermore, 8-OHdG is frequently used in studies to identify damage to DNA caused by oxidative stress [9, 10]. Sheep are reported to have high antioxidant capacity [12]. When menbutone (10 mg·kg<sup>-1</sup>) was administered to sheep, it can be stated that it will not cause oxidative stress and DNA damage and may be safe.

In the study, the effects of menbutone (10 mg·kg<sup>-1</sup>) administered to sheep on serum heart, liver, and kidney function parameters were investigated. It was determined that menbutone had no adverse effects on serum troponin I, CK-MB isoenzyme, ALP, ALT, AST, BUN, and creatinine levels in sheep ( $P>0.05$ , TABLE II). Troponin I and CK-MBiso levels are accepted as markers of heart damage and are preferred in studies investigating the cardiotoxic effects of drugs in sheep [17, 18].

Hepatic enzymes can be divided into two groups: markers of hepatocellular damage and markers of cholestasis. Increased serum ALT or AST indicates hepatocellular damage, while ALP levels elevate in bile duct damage [20]. Circulating creatinine and BUN concentrations are the most used markers of kidney function [19, 25].

It was reported that administration of menbutone at a dose of 400 mg·kg<sup>-1</sup> (IM) in rabbits resulted in increases in creatine kinase (CK), CK-MB, ALT, AST, and ALP levels. It was also stated that interstitial myocarditis and multifocal necrosis in the heart, necrosis with fatty infiltration in the liver, interstitial nephritis and renal tubular necrosis in the kidneys were determined [26]. Based on the findings of this research, it can be stated that menbutone at the recommended doses for sheep may not cause adverse effects on the heart, liver, and kidneys.

In the current research, the effect of menbutone on lipid metabolism parameters such as LDL and HDL levels in sheep was not determined ( $P>0.05$ ); however, it caused statistically significant fluctuations in cholesterol levels ( $P<0.05$ ) and a temporary increase in triglyceride level at 12<sup>th</sup> h ( $P<0.05$ ) (TABLE II). However, these changes were not dramatically significant and are likely within the reference range.

No clinically adverse effects were observed in the study when menbutone was administered to sheep. It has been stated that intravenous treatment of menbutone may cause side effects such as increased salivation, lacrimation, respiratory distress, tremors, restlessness, and involuntary urination and defecation [3].

It has been reported that menbutone might cause myonecrosis in a horse after accidental intramuscular injection [27] and the administration of metoclopramide and menbutone to a cat caused intestinal invagination [28]. When menbutone (10 mg·kg<sup>-1</sup>) was administered to Central Anatolian Merino sheep, it did not cause clinical side effects, and its use can be considered safe.

## CONCLUSIONS AND IMPLICATIONS

This study investigated the effects of menbutone administration at the recommended dose and duration on serum oxidative stress markers, cardiac, hepatic, and renal function, and lipid profile

parameters in sheep. According to the results of this study, it can be stated that menbutone has no effect on oxidative status parameters, does not affect heart, liver, and kidney functions, has a temporary minimal effect on lipid metabolism, and may be generally safe for sheep. However, studies are needed in target species with disease models.

## ACKNOWLEDGEMENTS

This study was sponsored by the Scientific Research Projects Coordination Office of Selcuk University (Project No: 25401155).

## Conflict of interest

The authors declare that they have any conflict of interest.

## BIBLIOGRAPHIC REFERENCES

- [1] European Medicines Agency (EMA). 16<sup>th</sup> Meeting of Committee for Veterinary Medicinal Products. [Internet]. The European Agency for the Evaluation of Medicinal Products. London (UK): European Medicines Agency. 1996 [cited 24 Mar 2025]. Available in: <https://goo.su/pEXDwn>
- [2] Tras B. Sindirim sistemi ilaçları [Digestive system drugs]. In: Yazar E, editor. Veteriner İlaç Rehberi ve Terapötik El Kitabı [Veterinary Drug Guide and Therapeutic Handbook]. Ankara (Türkiye): Nobel Medical Bookstores; 2024. p. 243–283. Turkish.
- [3] Hayvan Bilgi Sistemi-Tarbil [Animal Information System – Tarbil]. [Internet]. 2025 [cited 16 Mar 2025]. Turkish. Available in: <https://goo.su/4mMV6qQ>
- [4] Punzo A, Silla A, Fogacci F, Perillo M, Cicero AFG, Caliceti C. Bile acids and bilirubin role in oxidative stress and inflammation in cardiovascular diseases. *Diseases* [Internet]. 2024; 12(5):103. doi: <https://doi.org/qr9c>
- [5] Yazar E, Tras B. Serbest oksijen radikalleri, antioksidan enzimler ve antibiyotikler [Free oxygen radicals, antioxidant enzymes and antibiotics]. *J. Turk. Vet. Med. Assoc.* 2002; 14:42–44. Turkish.
- [6] Tabakoglu E, Durgut R. Veteriner hekimlikte oksidatif stres ve bazı önemli hastalıklarda oksidatif stresin etkileri [Oxidative stress in veterinary medicine and effects in some important diseases] *AVKAE Derg.* [Internet]. 2013 [cited 15 Mar 2025]; 3(1):69–75. Turkish. Available in: <https://goo.su/U3bzfHs>
- [7] Sezer K, Serbest oksijen radikallerinin hastalıkların patogeneziindeki rolü [Role of the free oxygen radicals on the pathogenesis of the diseases]. *FU. Sag. Bil. Vet. Derg.* [Internet]. 2014 [cited 12 Oct 2025]. 28(1):49–56. Turkish. Available in: <https://goo.su/UqSXW9s>
- [8] Calyniuk B, Grochowska-Niedworok E, Walkiewicz KW, Kawecka S, Popiołek E, Fatyga E. Malondialdehyde (MDA) – product of lipid peroxidation as marker of homeostasis disorders and aging. *Ann. Acad. Med. Siles.* [Internet]. 2016; 70:224–228. doi: <https://doi.org/g99sxz>

- [9] Di Minno A, Turnu L, Porro B, Squellerio I, Cavalca V, Tremoli E, Di Minno MN. 8-hydroxy-2-deoxyguanosine levels and cardiovascular disease: a systematic review and meta-analysis of the literature. *Antioxid. Redox Signal.* [Internet]. 2016; 24(10):548–555. doi: <https://doi.org/f8fx2h>
- [10] Pilger A, Rüdiger HW. 8-hydroxy-2-deoxyguanosine as a marker of oxidative DNA damage related to occupational and environmental exposures. *Int. Arch. Occup. Environ. Health.* [Internet]. 2006; 80:1–15. doi: <https://doi.org/b6r433>
- [11] Saracoglu A, Temel HE, Ergun B, Colak O. Oxidative stress-mediated cardiotoxicity of ciprofloxacin and ofloxacin in juvenile rats. *Drug Chem. Toxicol.* [Internet]. 2009; 32(3):238–242. doi: <https://doi.org/c2jzzx>
- [12] Coskun D, Parlak K, Dik B, Faki HE, Bahcivan E, Yazar E, Er A. Effect of enrofloxacin on the joint fluid/blood oxidative status and organ damage markers. *Annu. Res. Rev. Biol.* [Internet]. 2018; 25(3):1–7. doi: <https://doi.org/qr9d>
- [13] Coskun D, Canbar R, Korkmaz Y, Dik B, Er A, Yazar E. Determination of the effect of danofloxacin on 8-hydroxy-2-deoxyguanosine level. *Eurasian J. Vet. Sci.* [Internet]. 2019; 35(4):224–229. doi: <https://doi.org/qr9f>
- [14] Yazar E. *Veteriner İlaç ve Aşı – A dan Z ye (2018 – 2019)* [Veterinary Drugs and Vaccines – A to Z (2018 – 2019)]. Ankara (Türkiye): Nobel Medical Bookstores; 2018. 306 p. Turkish.
- [15] Kaya S, Ünsal A. İlaçların istenmeyen etkileri [Adverse effects of medications]. In: Kaya S, Pirinççi İ, Bilgili A, editors. *Veteriner Hekimliğinde Farmakoloji* [Veterinary Applied Pharmacology]. 3<sup>rd</sup> ed. Ankara (Türkiye): Medisan; 2002. p.142–152.
- [16] Dogan F. Pharmacovigilance in veterinary profession. *Eurasian J. Vet. Sci.* [Internet]. 2011 [cited 15 Mar 2025]; 27(1):19–25. Available in: <https://goo.su/qkWms>
- [17] Corum O, Er A, Dik B, Eser H, Bahcivan E, Yazar E. Determination of the safety of tulathromycin in sheep. *Eurasian J. Vet. Sci.* [Internet]. 2015; 31(3):152–157. doi: <https://doi.org/qr9g>
- [18] Corum O, Dik B, Bahcivan E, Eser H, Er A, Yazar E. Cardiac safety of gamithromycin in ewes. *Eurasian J. Vet. Sci.* [Internet]. 2016; 32(4):242–245. doi: <https://doi.org/qr9h>
- [19] Kerr MG. *Veterinary Laboratory Medicine. Clinical biochemistry and haematology.* 2<sup>nd</sup> ed. London (UK): Blackwell Science; 2002. 386 p. Available in: <https://goo.su/ozVrL>
- [20] Turgut K. Karaciğer testi (Liver test). In: Turgut K, editor. *Veteriner Klinik Laboratuvar Teşhis* [Veterinary Clinical Laboratory Diagnostics]. Konya (Türkiye): Gardeners Printing House; 2000. p. 202–257.
- [21] Turgut K. Endokrin, metabolik ve lipid bozukluklarının testleri [Tests for endocrine, metabolic, and lipid disorders]. In: Turgut K, editor. *Veteriner Klinik Laboratuvar Teşhis* [Veterinary Clinical Laboratory Diagnostics]. Konya (Türkiye): Gardeners Printing House; 2000. p. 416–486.
- [22] Er A, Corum O, Eser H, Bahcivan E, Dik B, Yazar E. Effect of dexamethasone treatment on blood oxidative status and prostaglandin F<sub>2α</sub> metabolite levels in ram. *Eurasian J. Vet. Sci.* [Internet]. 2016; 32(2):89–93. doi: <https://doi.org/qsbs>
- [23] Ider M, Naseri A, Parlak TM, Zhunushova A, Yazar E. Safety of an antiprotozoal drug combination in sheep. *Eurasian J. Vet. Sci.* [Internet]. 2020; 36(2):115–120. doi: <https://doi.org/qsbt>
- [24] Canbar R, Uslu M, Arslan MS, Yazar E. Effect of combined application of ivermectin and praziquantel on oxidative stress and selected biochemical parameters in sheep. *Eurasian J. Vet. Sci.* [Internet]. 2023; 39(1):25–29. doi: <https://doi.org/qsbv>
- [25] Cobrin AR, Blois SL, Kruth SA, Abrams-Ogg ACG, Dewey C. Biomarkers in the assessment of acute and chronic kidney diseases in the dog and cat. *J. Small Anim. Pract.* [Internet]. 2013; 54(12):647–655. doi: <https://doi.org/f5htzf>
- [26] El Okle SO, Tohamy GH, Lebda AM. Evaluation of acute toxicity of genabilic acid (menbutone 10%) in rabbits. *World Rabbit Sci.* [Internet]. 2014; 22(3):215–222. doi: <https://doi.org/qsbw>
- [27] Slowikowska M, Siwinska N, Zak A, Borowicz H, Kubiak K, Niedzwiedz A. Myonecrosis in a horse after an intramuscular injection of menbutone. *Med. Weter.* [Internet]. 2018; 74(12):795–798. doi: <https://doi.org/qsbx>
- [28] Lukanc B, Pogorevc E, Kastelic A, Erjavec V. Retrograde jejunal intussusception in a one-year-old cat after treatment with metoclopramide and menbutone. *Slov. Vet. Res.* [Internet]. 2014 [cited 15 Mar 2025]; 51(4):201–207. Available in: <https://goo.su/iFQhUFV>